# **Novel Approaches to Spectral Processing and Quantification**

Sarah J. Nelson, PhD

Program in Bioengineering and Department of Radiology, UCSF

Magnetic Resonance Spectroscopy (MRS) is increasingly being used as an adjunct to conventional imaging to provide information concerning the viability, proliferative status and functional behavior of living tissue for basic science, pre-clinical and clinical situations. Although the procedures for acquiring 1-H MRS data on animal and human scanners are fairly common, there are still no commercial standards for post-processing and quantitative analysis. This limits the ability to compare results across platforms and means that many studies are still either using qualitative analyses or taking their results offline for post-processing. A further complication for the interpretation of MRS data is the trend towards using multi-channel radiofrequency coils. While there are numerous possibilities for combining the contributions from different elements using parallel imaging reconstruction methods, there have been a relatively small number of applications to MRS data. This presentation will summarize the state of the art in reconstruction, analysis and interpretation of spectral data with an emphasis on novel approaches and future needs.

# H-1 MR Spectroscopy of the Brain a 1.5T

The use of water suppressed H-1 spectroscopy has been facilitated by the availability of pulse sequences for PRESS and STEAM volume selection that can be used routinely in the brain in single or multi-voxel mode. While the data quality in regions that avoid the sinuses is generally good there are a number of artifacts that arise due to lipid contamination, motion and gradient imperfections. In a recent review, Kreis (1) provided an extensive description of the common spectral artifacts that can be observed with single voxel and multi-voxel spectral acquisitions in the brain. It is important that these artifacts become more widely understood so that the clinical interpretation of the data is accurate. Analysis of STEAM and PRESS datasets from the brain are typically performed using automated software packages (2-5). While model-based functions are typically such as LCModel and AMARES are typically used for fitting short echo time spectra (6-7), there are still challenges for short echo times in terms of managing the baseline components that arise due to the presence of macromolecules (8).

Although an increasing number of researchers are using echo planar or spiral k-space sampling approaches, these are not yet available for routine applications. These options are necessary for obtaining increased coverage and large 3-D matrix sizes within a relatively modest acquisition time but lead to challenges in terms of maintaining data quality and developing more sophisticated methods for quantification, reconstruction and post-processing. Ebel et al (9) showed that although automated processing of such data was possible and that a large region of the brain could be covered there were a relatively large percentage of voxels for which there was poor linewidth or significant lipid contamination. Other alternative k-space sampling strategies and methods for reducing lipid components and have been proposed recently, including changes in acquisition parameters and post-processing routines (10-14).

### H-1 MRS at higher field strengths and from other organs

The increasing availability of clinical 3T MR scanners has provided spectra with higher signal to noise ratio from the brain (15-19). The T2 relaxation times of brain metabolites are

longer at 3T than at 1.5T and so the actual increase in signal to noise ratio is less than a factor of two but there is typically improved spectral resolution (16), which leads to more reliable quantification of metabolites such as myo-inositol and glutamate using approaches such as J-resolved MRS with LCModel fitting (17-19).

While the brain is the most common region being studied with H-1 MRS, there are a substantial number of applications in the prostate at (20-22). Quantification of prostate spectra requires estimation of levels of choline, creatine and citrate. The parameter most commonly used to infer tumor versus normal or benign tissue is the ratio of (choline+creatine)/citrate. The majority of the studies to date have used frequency domain integration of peak regions as opposed to time domain fitting to obtain these values. A recent comparison of the two techniques showed that at 1.5T the accuracy of time domain fitting was improved for well shimmed spectra with high signal to noise ratio but that the simpler frequency domain approach was more robust, especially in the presence of the broad polyamine resonance in normal prostate tissue (23). The fitting of prostate spectral peaks is more challenging at 3T due to the strong coupling of the citrate resonances and requires either a change in acquisition parameters or more complex modeling of the spectral components (24-27).

There have also been an increasing number of applications of MRS to the breast at 1.5T and 4T (28-35). In this case the main resonance being observed is choline and the challenge is to suppress large lipid peaks and obtain at least a relative measure of peak intensity. Bolan et al have addressed this at 4T in a series of papers that have come up with a robust strategy for acquisition and quantification (31-34). Together with the work of Katz-Bull et al (29), these have laid the groundwork for more extensive clinical studies. Similar challenges are faced in other body applications. Preliminary studies in this regard have included liver, thorax, muscle, head and neck, and cervix (36-38).

### **Multi-channel H-1 MRS**

The availability of MR scanners with multiple receive channels has expanded the usage of phased array coils and raised the question of how to best combined the signals from individual elements. As reviewed in the original papers (39-40), a variety of different approaches can be taken, depending on whether the goal is to provide phase sensitive spectra and to correct for spatial variations in intensity of the spectra due to the non-uniform rf profiles of individual coil elements (41-43). The latter requires estimates of the profiles of individual coils which can be obtained from theoretical estimates, empirical measurements in phantoms, by using residual water as a reference or by filtering of in vivo images that have been acquired with minimal (usually proton density weighted) contrast. For phase sensitive spectra it may be necessary to process each channel separately and then combine the results afterwards. This has been shown to be feasible but does require increased processing time.

The trend in MR imaging towards more rapid, multichannel acquisitions has provided a revolution in the use of so-called parallel reconstruction methods which make use of the fact that there is distinct spatial information in the individual coil elements. This can be used to reconstruct data with a higher resolution than would have been possible by simply taking the fourier transform of the original phase encoded arrays. This is particularly relevant at higher field strengths, where the increased signal to nose ratio means that larger acquisition matrices and/or faster scan times are feasible. Because MRSI uses phase encoding or other types of k-space sampling in multiple directions, it can benefit from speed-up factors in more than one dimension. Examples of the use of the SENSE were presented by Dydak et al (44-45). There are clearly

trade-offs in terns of spectral artifacts and reconstruction times that must be considered in using such parallel approaches (46). These need to be studied further before these techniques are applied in a routine clinical setting.

#### **Multinuclear MRS**

The increased availability of MR scanners with in higher field strength has meant that there is beginning to be a resurgence of the applications of multi-nuclear MRS with a particular emphasis on P-31 (47) and C-13 (48-51) MRS. There have been relatively few new insights into the processing of such data beyond the use of fitting model functions with prior information, but this is likely to change as the number of studies being performed increases (52). A further motivation has been the promise of the availability of hyperpolarized C-13 tracers which is providing challenges in terms of having very high signal to noise ratio but requiring very rapid acquisitions (53). This field is just in its infancy but is of major interest for researchers engaged in developing data processing algorithms as it will need novel k-space sampling strategies, parallel reconstruction methods, time domain fitting of truncated signals and dynamic modeling of signal decay.

# Tissue Classification by MRI and MRS

With the availability of a larger body of clinical MRS data there has been the need to apply multi-variate statistical techniques and artificial intelligence approaches to determining which metabolite peaks and which combination of imaging methods provide the most relevant information. The largest number of these has been to the analysis of data from brain tumors and has included consideration of the MRS data alone (54-59) or in conjunction with conventional MRI, perfusion MRI or diffusion MRI (60-65). The consensus is that high choline and low n-acetylaspartate distinguish tumor from normal tissue and that high lactate and lipid or low creatine are indicative of increased malignancy. This may be reflected in higher histological grade and/or worse outcome (61-65). Similar strategies are being applied to conventional MRI, dynamic contrast uptake imaging and MRS data from patients with prostate cancer and have shown that the MRS data definitely add to the diagnosis and ability to assess response to therapy (66-68). Methods such as canonical correlation analysis and neural networks are likely to play an increasing role in determining the clinical benefits of new MRI and MRS techniques (69-71).

### References

- 1. Kreis, R. Issues of spectral quality in clinical 1-H magnetic Resonance Spectroscopy and a gallery of artifacts, NMR in Biomed. 17: 361-381, 2004.
- 2. Coron A, Vanhamme L., Antione J-P. Van Hecke P. and Van Huffel S. The Filtering Approach to Solvent Peak Suppression in MRS: A Critical Review, J. Mag.Res (2001) 152:26-40.
- 3. Nelson SJ.Analysis of volume MRI and MR spectroscopic imaging data for the evaluation of patients with brain tumors. Magn Reson Med. 2001, 46:228-39.
- 4. Van Huffel S, Wang Y, Vanhamme L, Van Hecke P. Automatic frequency alignment and quantitation of single resonances in multiple magnetic resonance spectra via complex principal component analysis. J Magn Reson. 2002, 158:1-14.
- 5. Elster C, Schubert F, Link A, Walzel M, Seifert F, Rinneberg H. Quantitative magnetic resonance spectroscopy: semi-parametric modeling and determination of uncertainties. Magn Reson Med. 2005, 53:1288-96.

- 6. Soher B, Maudesley A. Evaluation of Variable Line-shape Models and Prior Information in automated 1-H spectroscopic imaging analysis, Mag Res Med 52: 1246-1254, 2005.
- 7. Kanowski M., Kaufmann J., Braun J., Bernarding J., and Templelmann C., Quantification of Simulated Short Echo Time 1-H Brain Spectra by LC Model and AMARES. Mag, Res. Med, 51: 904-912, 2004
- 8. Hofmann L, Slotboom J., Boesch C. and Kreis R. Characterization of the Macromolecular Baseline in Localized 1h\_MR Spectra of Human Brain, Mag. Res. Med. 46: 855-863, 2001
- 9. Ebel A., Soher B., Maudesley A, Assessment of 3D Proton MR Echo-Planar Spectroscopic Imaging using Automated Spectral Analysis, Mag, Res. Med, 46: 1072-1078, 2001.
- 10. Cunningham CH, Vigneron DB, Chen AP, Xu D, Nelson SJ, Hurd RE, Kelley DA, Pauly JM. Design of flyback echo-planar readout gradients for magnetic resonance spectroscopic imaging. Magn Reson Med. 2005 54(5):1286-9.
- 11. Serrai H., Senhadjii L., Wang G, Akoka S, Stroman P, Mag.Res.Med. Lactate Doublet Quantification and Lipid Signal Suppression using a new bioexponential decay filter: Application to simulated and 1-H MRS Brain Tumor Time Domain Data 50: 6230626, 2003.
- 12. Sarkar S, Heberlein K, Hu X. Truncation artifact reduction in spectroscopic imaging using a dual-density spiral k-space trajectory. Magn Reson Imaging. 2002 20(10):743-57.
- 13. Tran TK, Vigneron DB, Sailasuta N, Tropp J, Le Roux P, Kurhanewicz J, Nelson S, Hurd R.Very selective suppression pulses for clinical MRSI studies of brain and prostate cancer. Magn Reson Med. 2000 43(1):23-33.
- 14. Star-Lack JM, Spielman DM. Zero-quantum filter offering single-shot lipid suppression and simultaneous detection of lactate, choline, and creatine resonances. Magn Reson Med. 2001 46(6):1233-7.
- 15. Barker PB, Hearshen DO, Boska MD. Single-voxel proton MRS of the human brain at 1.5T and 3.0T. Magn Reson Med. 2001 May;45(5):765-9.
- 16. Li B.S., Regal J. and Gonen O. SNR versus Resolution in 3D 1-H MRS of the Human Brain at High Magnetic Fields, Mag. Res. Med. 46:1049-1053, 2001.
- 17. Srinivasan R, Vigneron D, Sailasuta N, Hurd R, Nelson S.A comparative study of myoinositol quantification using LCmodel at 1.5 T and 3.0 T with 3 D 1H proton spectroscopic imaging of the human brain.Magn Reson Imaging. 2004 22(4):523-8.
- 18. Hurd R, Sailasuta N, Srinivasan R, Vigneron DB, Pelletier D, Nelson SJ. Measurement of brain glutamate using TE-averaged PRESS at 3T. Magn Reson Med. 2004 Mar;51(3):435-40.
- 19. Mayer D, Spielman DM. Detection of glutamate in the human brain at 3 T using optimized constant time point resolved spectroscopy. Magn Reson Med. 2005 Aug;54(2):439-42.
- 20. Jung JA, Coakley FV, Vigneron DB, Swanson MG, Qayyum A, Weinberg V, Jones KD, Carroll PR, Kurhanewicz J. Prostate depiction at endorectal MR spectroscopic imaging: investigation of a standardized evaluation system. Radiology. 2004, 233:701-8.
- 21. Wang L, Mullerad M, Chen HN, Eberhardt SC, Kattan MW, Scardino PT, Hricak H. Prostate cancer: incremental value of endorectal MR imaging findings for prediction of extracapsular extension. Radiology. 2004, 232:133-9.
- 22. Barnes AS, Haker SJ, Mulkern RV, So M, D'Amico AV, Tempany CM. Magnetic resonance spectroscopy-guided transperineal prostate biopsy and brachytherapy for recurrent prostate cancer. Urology. 2005 66(6):1319.
- 23. Pels P, Ozturk E, Swanson M, VanHamme L. Kurhanewicz J, Nelson SJ, Van Huffel S., Quantification of Prostate MRSI Data by Model-Based Time Domain Fitting and Frequency Domain Analysis, NMR in Biomed., in press 2005.

- 24. Futterer JJ, Scheenen TW, Huisman HJ, Klomp DW, van Dorsten FA, Hulsbergen-van de Kaa CA, Witjes JA, Heerschap A, Barentsz JO. Initial experience of 3 tesla endorectal coil magnetic resonance imaging and 1H-spectroscopic imaging of the prostate. Invest Radiol. 2004 39(11):671-80.
- 25. Cunningham CH, Vigneron DB, Marjanska M, Chen AP, Xu D, Hurd RE, Kurhanewicz J, Garwood M, Pauly JM. Sequence design for magnetic resonance spectroscopic imaging of prostate cancer at 3 T.Magn Reson Med. 2005 53(5):1033-9.
- 26. Kim DH, Margolis D, Xing L, Daniel B, Spielman D. In vivo prostate magnetic resonance spectroscopic imaging using two-dimensional J-resolved PRESS at 3 T. Magn Reson Med. 2005 53(5):1177-82.
- 27. Trabesinger AH, Meier D, Dydak U, Lamerichs R, Boesiger P. Optimizing PRESS localized citrate detection at 3 Tesla. Magn Reson Med. 2005 54(1):51-8
- 28. Bakken IJ, Gribbestad IS, Singstad TE, Kvistad KA. External standard method for the in vivo quantification of choline-containing compounds in breast tumors by proton MR spectroscopy at 1.5 Tesla. Magn Reson Med. 2001 46(1):189-92.
- 29. Katz-Brull R, Lavin PT, Lenkinski RE. Clinical utility of proton magnetic resonance spectroscopy in characterizing breast lesions. J Natl Cancer Inst. 2002 21;94(16):1197-203.
- 30. Kim JK, Park SH, Lee HM, Lee YH, Sung NK, Chung DS, Kim OD. In vivo 1H-MRS evaluation of malignant and benign breast diseases. Breast. 2003 12(3):179-82.
- 31. Bolan PJ, DelaBarre L, Baker EH, Merkle H, Everson LI, Yee D, Garwood M. Eliminating spurious lipid sidebands in 1H MRS of breast lesions. Magn Reson Med. 2002 48(2):215-22.
- 32. Bolan PJ, Meisamy S, Baker EH, Lin J, Emory T, Nelson M, Everson LI, Yee D, Garwood M. In vivo quantification of choline compounds in the breast with 1H MR spectroscopy. Magn Reson Med. 2003 50(6):1134-43.
- 33. Bolan PJ, Henry PG, Baker EH, Meisamy S, Garwood M. Measurement and correction of respiration-induced B0 variations in breast 1H MRS at 4 Tesla. Magn Reson Med. 2004 52(6):1239-45
- 34. Bolan PJ, Nelson MT, Yee D, Garwood M. Imaging in breast cancer: Magnetic resonance spectroscopy. Breast Cancer Res. 2005;7(4):149-52. Epub 2005.
- 35. Jacobs MA, Barker PB, Argani P, Ouwerkerk R, Bhujwalla ZM, Bluemke DA. Combined dynamic contrast enhanced breast MR and proton spectroscopic imaging: a feasibility study. J Magn Reson Imaging. 2005 21(1):23-8.
- 36. Li C-W, Kuo Y.C. Chen C-Y, Kuo Y-T, Chuo Y-Y, She F-O, Liu G-C. Quantification of Choline in Human Hepatic Tumors by Proton MR Spectroscopy at 3T Mag.Res.Med 53: 770-776,2005
- 37. Katz-Brull R, Rofsky NM, Lenkinski RE. Breathhold abdominal and thoracic proton MR spectroscopy at 3T.Magn Reson Med. 2003 Sep;50(3):461-7Mahon MM, Williams AD, Soutter WP, Cox IJ, McIndoe GA, Coutts GA, Dina R, deSouza NM. 1H magnetic resonance spectroscopy of invasive cervical cancer: an in vivo study with ex vivo corroboration. NMR Biomed. 2004, 17:1-9.
- 38. Seenu V, Pavan Kumar MN, Sharma U, Gupta SD, Mehta SN, Jagannathan NR. Potential of magnetic resonance spectroscopy to detect metastasis in axillary lymph nodes in breast cancer. Magn Reson Imaging. 2005 23(10):1005-10. Epub 2005
- 39. Wald LL, Moyher SE, Day MR, Nelson SJ, Vigneron DB.Proton spectroscopic imaging of the human brain using phased array detectors. Magn Reson Med. 1995 34(3):440-5.
- 40. Wright SM, Wald LL. Theory and application of array coils in MR spectroscopy. NMR

- Biomed. 1997 10(8):394-410.
- 41. Mark A. Brown, Time Domain Combination of MR spectroscopy data acquired using phased array coils.Mag. Res. Med. 52: 1207-1214, 2005
- 42. Sandgren N, Stoica P, Frigo FJ, Selen Y. Spectral analysis of multichannel MRS data. J Magn Reson. 2005 175(1):79-91
- 43. Prock T, Collins DJ, Dzik-Jurasz AS, Leach MO. An algorithm for the optimum combination of data from arbitrary magnetic resonance phased array probes. Phys Med Biol. 2002 Jan 21;47(2):N39-46
- 44. Dydak U, Weiger M, Pruessmann KP, Meier D, Boesiger P. Sensitivity-encoded spectroscopic imaging. Magn Reson Med. 2001 46(4):713-22.
- 45. Dydak U, Pruessmann KP, Weiger M, Tsao J, Meier D, Boesiger P. Parallel spectroscopic imaging with spin-echo trains. Magn Reson Med. 2003 50(1):196-200.
- 46. Zhao X, Prost RW, Li Z, Li SJ. Reduction of artifacts by optimization of the sensitivity map in sensitivity-encoded spectroscopic imaging. Magn Reson Med. 2005 53(1):30-4.
- 47. Mattei JP, Bendahan D, Cozzone P. P-31 Magnetic resonance spectroscopy. A tool for diagnostic purposes and pathophysiological insights in muscle diseases. Reumatism. 2004 56(1):9-14.
- 48. Morris P and Bachelard H, Reflections on the application of C-13 MRS to research on brain metabolism, NMR in Biomed 16:303-312, 2003.
- 49. De Graaf R.A., Mason G., Patel A.B., Behar K.L. and Rothman D.L. In vivo 1H-C-13 NMR spectroscopy of cerebral metabolism NMR Biomed 2003: 16: 339-357.
- 50. Gruetter R, Adriany G, Choi in-Young, Henry P-G, Lei H, Oz Gulin, Localized in vivo C-13 NMR spectroscopy of the brain NMR in Biomed. 2003 16: 313-338.
- 51. Ross B, Lin A, Harris K, Bhattacharya P, Schweinsburg B. Clinical experience with 13C MRS in vivo. NMR Biomed. 2003 16(6-7):358-69.
- 52. Shic F, Ross B. Automated data processing of [1H-decoupled] 13C MR spectra acquired from human brain in vivo. J Magn Reson. 2003 Jun;162(2):259-68.
- 53. Bhattacharya P, Harris K, Lin AP, Mansson M, Norton VA, Perman WH, Weitekamp DP, Ross BD.Ultra-fast three dimensional imaging of hyperpolarized (13)C in vivo. MAGMA. 2005 Oct;18(5):245-56. Epub 2005 Nov 23.
- 54. McKnight TR, Noworolski SM, Vigneron DB, Nelson SJ. An automated technique for the quantitative assessment of 3D-MRSI data from patients with glioma. J Magn Reson Imaging. 2001 Feb;13(2):167-77.
- 55. Lukas L, Devos A, Suykens JA, Vanhamme L, Howe FA, Majos C, Moreno-Torres A, Van der Graaf M, Tate AR, Arus C, Van Huffel S. Brain tumor classification based on long echo proton MRS signals. Artif Intell Med. 2004, 31:73-89.
- 56. Devos A, Lukas L, Suykens JA, Vanhamme L, Tate AR, Howe FA, Majos C, Moreno-Torres A, van der Graaf M, Arus C, Van Huffel S. Classification of brain tumours using short echo time 1H MR spectra. J Magn Reson. 2004, 170:164-75.
- 57. Majos C, Julia-Sape M, Alonso J, Serrallonga M, Aguilera C, Acebes JJ, Arus C, Gili J. Brain tumor classification by proton MR spectroscopy: comparison of diagnostic accuracy at short and long TE. AJNR Am J Neuroradiol. 2004, 25:1696-704.
- 58. Tate AR, Majos C, Moreno A, Howe FA, Griffiths JR, Arus C. Automated classification of short echo time in vivo 1H brain tumor spectra: a multicenter study. Magn Reson Med. 2003, 49:29-36.
- 59. Underwood J, Tate AR, Luckin R, Majos C, Capdevila A, Howe F, Griffiths J, Arus C. A

- prototype decision support system for MR spectroscopy-assisted diagnosis of brain tumours Medinfo. 2001, 10:561-5.
- 60. Simonetti AW, Melssen WJ, Szabo de Edelenyi F, van Asten JJ, Heerschap A, Buydens LM. Combination of feature-reduced MR spectroscopic and MR imaging data for improved brain tumor classification. NMR Biomed. 2005, 18:34-43.
- 61. Devos A, Simonetti AW, van der Graaf M, Lukas L, Suykens JA, Vanhamme L, Buydens LM, Heerschap A, Van Huffel S. The use of multivariate MR imaging intensities versus metabolic data from MR spectroscopic imaging for brain tumour classification. J Magn Reson. 2005, 173:218-28.
- 62. Kuznetsov YE, Caramanos Z, Antel SB, Preul MC, Leblanc R, Villemure JG, Pokrupa R, Olivier A, Sadikot A, Arnold DL. Proton magnetic resonance spectroscopic imaging can predict length of survival in patients with supratentorial gliomas. Neurosurgery. 2003, 53:565-74; discussion 574-6.
- 63. Li X, Jin H, Lu Y, Oh J, Chang S, Nelson SJ. Identification of MRI and 1H MRSI parameters that may predict survival for patients with malignant gliomas. NMR Biomed. 2004, 17:10-20.
- 64. Law M, Yang S, Wang H, Babb JS, Johnson G, Cha S, Knopp EA, Zagzag D. Glioma grading: sensitivity, specificity, and predictive values of perfusion MR imaging and proton MR spectroscopic imaging compared with conventional MR imaging. AJNR Am J Neuroradiol. 2003, 24:1989-98.
- 65. Chiang IC, Kuo YT, Lu CY, Yeung KW, Lin WC, Sheu FO, Liu GC. Distinction between high-grade gliomas and solitary metastases using peritumoral 3-T magnetic resonance spectroscopy, diffusion, and perfusion imagings. Neuroradiology. 2004 46:619-27.
- 66. van Dorsten FA, van der Graaf M, Engelbrecht MR, van Leenders GJ, Verhofstad A, Rijpkema M, de la Rosette JJ, Barentsz JO, Heerschap A. Combined quantitative dynamic contrast-enhanced MR imaging and (1)H MR spectroscopic imaging of human prostate cancer. J Magn Reson Imaging. 2004, 20:279-87.
- 67. Noworolski SM, Henry RG, Vigneron DB, Kurhanewicz J. Dynamic contrast-enhanced MRI in normal and abnormal prostate tissues as defined by biopsy, MRI, and 3D MRSI Magn Reson Med. 2005 53(2):249-55.
- 68. Pucar D, Koutcher JA, Shah A, Dyke JP, Schwartz L, Thaler H, Kurhanewicz J, Scardino PT, Kelly WK, Hricak H, Zakian KL.Preliminary assessment of magnetic resonance spectroscopic imaging in predicting treatment outcome in patients with prostate cancer at high risk for relapse. Clin Prostate Cancer. 2004 3(3):174-81
- 69. Hasumi M, Suzuki K, Taketomi A, Matsui H, Yamamoto T, Ito K, Kurokawa K, Aoki J, Endo K, Yamanaka H. The combination of multi-voxel MR spectroscopy with MR imaging improve the diagnostic accuracy for localization of prostate cancer. Anticancer Res. 2003;23(5b):4223-7.
- 70. Laudadio T, Pels P, De Lathauwer L, Van Hecke P, Van Huffel S. Tissue segmentation and classification of MRSI data using canonical correlation analysis. Magn Reson Med. 2005, 54:1519-29.
- 71. Laudadio T, Pels P, De Lathauwer L, Van Hecke P, Van Huffel S. Tissue segmentation and classification of MRSI data using canonical correlation analysis. Magn Reson Med. 2005 54(6):1519-29